

# New Solvent Grade Targeted for Trace Analysis by UHPLC-MS

Subhra Bhattacharya, Deva H. Puranam, and Stephen C. Roemer

Thermo Fisher Scientific – Fisher Chemical, One Reagent Lane, Fair Lawn, NJ





Fisher Chemical has developed a new solvent grade, UHPLC-MS Optima<sup>TM</sup>, for mobile phases targeting trace analysis by UHPLC-MS. These ultra-pure solvents will provide a very low mass noise level in both positive and negative mode ionization, minimal metal ion content, and very low UHPLC-UV response using photo diode array detection. Fisher Chemical's high purity solvents are specifically qualified for UHPLC-MS and offered in Acetonitrile, Methanol, and Water.

Solvent	Pack size	Packaging	<b>Catalog Number</b>
Acetonitrile	1L	Borosilicate Glass	A956-1
Methanol	1L	Borosilicate Glass	A458-1
Water	1L	Borosilicate Glass	W8-1

#### Introduction

Ultra-high performance liquid chromatography (UHPLC) is associated with submicron particle size column and high pressure flow resulting in increased resolution and sensitivity for complex sample mixtures and increased speed of analysis. Mass spectrometry (MS) enables the detection and identification of analytes at the parts per trillion level. UHPLC coupled with MS (UHPLC-MS) is a powerful tool in analytical chemistry that requires mobile phase solutions prepared with exceptionally pure solvents permitting trace analysis. UHPLC-MS Optima<sup>TM</sup>, the new Fisher Chemical solvent grade for mobile phases targeting UHPLC-MS will show a very low mass noise level in both positive and negative mode ionization, minimal metal ion content, and very low UHPLC-UV response using photodiode array detection.

# **Material and Methods**

- Mobile phase: acetonitrile (ACN), methanol, and water were evaluated; all three solvents are from a new grade, Optima® UHPLC-MS, that will provide very low mass noise level in both positive and negative mode ionization.
- Instrument: Thermo Scientific Accela UHPLC system comprised of an auto-sampler, photodiode array detector, and attached to an LTQ-XL mass spectrometer equipped with an electro-spray ionization interface.
- Column: Thermo Scientific Hypersil GoldTM column (50 mm x 2.1 mm, 1.9 micron), Catalog No: 26-102-052130.
- Standards: propazine (SPEX CertiPrep, S-3170,1000 μg/mL) as the positive mode standard and chloramphenicol (SPEX CertiPrep, S-4032, 1000 μg/mL) as negative mode standard. These standards provided adequate ionization without any additive applied to the mobile phase.
- The mass spectrometer was operated in full scan ESI-MS from 100 to 1500 amu. The collision induced dissociation (CID) mass spectra were obtained with helium as the collision gas after isolation of the particular precursor ion. Other parameters including gas flow and capillary voltage were adjusted as required.

#### • HPLC Gradient:

• 0 — 0.5 min: 90% water, 10% ACN

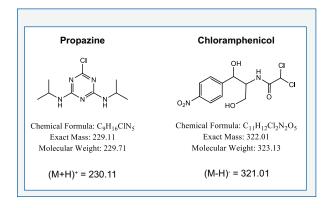
• 0.5 min — 2.0 min: 0% water, 100% ACN

• 2 min — 5 min: 100% ACN

• Post run 5.1 min — 10 min: 90% water, 10% ACN

Flow rate: 0.6 mL/min for water/ACN, 0.5 mL/min for water/methanol

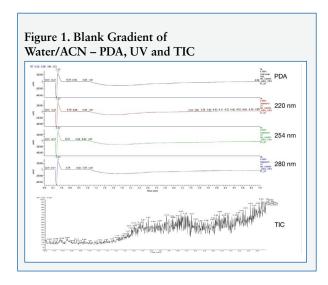
• Injection volume: 5 µl

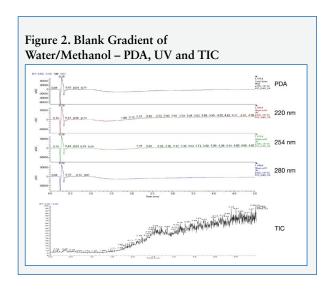


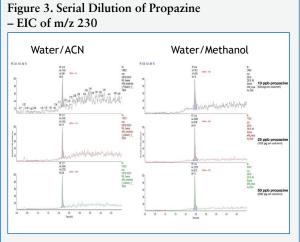


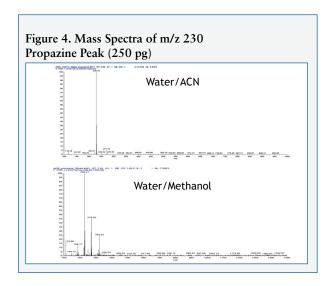
# **Results**

- Mobile phase solvent purity was evaluated by linking UHPLC-MS sensitivity to trace analysis of positive and negative mode standards (Figs. 1–10).
- Propazine was used as positive mode standard (Figs. 3–6) and chloramphenicol as negative mode standard (Figs. 7–10) in order to assess interfering baseline peaks in both full scan ESI-MS and CID generated product ions.









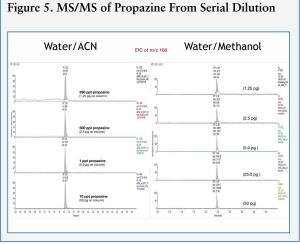




Figure 6. MS/MS Spectra of Propazine in Two
Different Solvent Systems

Water/ACN

Water/ACN

Fragmentation of Propazine

Chemical Formula C, III, Chemical Formula C, III,

Water/ACN

Elc m/z 321

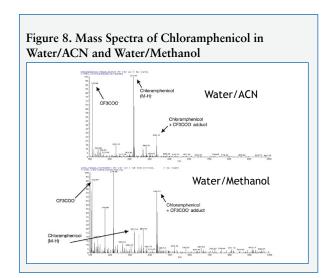
Water/Methanol

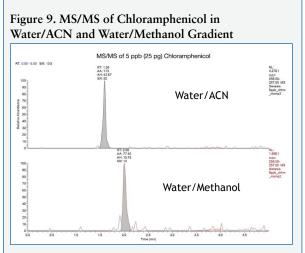
Figure 7. Serial Dilution of Chloramphenicol

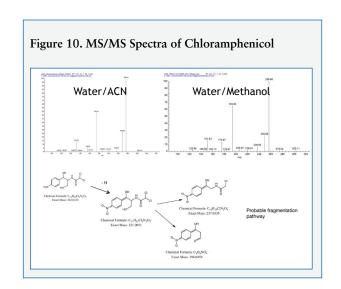
Water/Methanol

Figure 8. Serial Dilution of Chloramphenicol

Figure







# **Discussion**

- Blank gradient of water/ACN and water/methanol in PDA, UV and mass spectra is shown in Figs. 1 and 2. No extraneous peaks were observed in either solvent system.
- Trace level of propazine (10 ppb = 10 pg/μL) was detected in positive mode ionization both in water/ACN and water/methanol gradient by EIC after full scale data acquisition (100 1500 amu). The signal to noise ratio of the peak was observed below the level of quantitation (LOQ) in the ACN gradient. However, a 25 pg/μL concentration (25 ppb) of propazine showed S/N >10 in both solvent systems (Fig. 3).
- MS/MS of propazine peak (m/z 230) was accomplished using 250 fg/µL (250 ppt) of analyte, and the signal to noise ratio of the m/z 188 product ion was monitored. For both solvent systems, the propazine product ion showed S/N >30 (Figs. 5 and 6) which is consistent with the detection sensitivity of the LTQ-XL ion trap mass spectrometer.
- In negative mode ionization, 50 pg/ $\mu$ L (50 ppb) of chloramphenicol showed S/N >10 in the ACN and methanol gradients (Fig. 7). Both solvent systems showed m/z 435 due to TFA (m/z 113) adduct formation coming from the system (Fig. 8).
- MS/MS of 5 pg/μl (5 ppb) chloramphenicol generated the product ion of m/z 257 with a S/N >10 (Figs. 9 and 10).

## **Conclusions**

- Using the new Optima® UHPLC-MS mobile phase solvents, trace amounts of propazine and chloramphenicol standards showed significant peak height without interfering background peaks in both full scan ESI-MS and CID generated product ions.
- Monitoring the CID generated product ion peak is common practice for evaluating MS sensitivity, and we have developed a similar approach to assess the purity of mobile phase solvents.
- Propazine (m/z 230) generated the product ion peak of m/z 188 with a signal to noise ratio >30 at 250 ppt using a water/acetonitrile mobile phase.





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ENA 23, Zone 1, nr 1350 Janssen Pharmaceuticalaan 3a 2440 Geel Belgium www.acros.com